## Poster 59. Haplotype analysis of the leaf stripe resistance locus Rdg2a in different barley genotypes.

C. Biselli <sup>1,2</sup>, S. Urso <sup>1</sup>, N. Stein <sup>3</sup>, L. Cattivelli <sup>1</sup>, and G. Valè <sup>1,2</sup>.

<sup>1</sup> Genomic Research Center, CRA–GPG, Via S. Protaso 302, I-29017 Fiorenzuola d'Arda (PC), Italy; <sup>2</sup> Unità di ricerca per la risicoltura, CRA–RIS, Strada Statale 11 per Torino km 2,5, 13100, Vercelli, Italy; and <sup>3</sup> Leibniz Institute of Plant Genetics and Crop Plant Research, 06466 Gatersleben, Germany.

Leaf stripe disease on barley is caused by the seed-transmitted hemi-biotrophic fungus Pyrenophora graminea (anamorph Drechslera graminea). Race-specific resistance to leaf stripe is controlled by two known Rdg (Resistance to Drechslera graminea) genes, the H. spontaneum-derived Rdg1a, mapped to chromosome 2HL, and Rdg2a, identified in H. vulgare and mapped on chromosome 7HS and cloned in the resistant cultivar Thibaut. The Rdg2a locus contains a gene cluster of three sequence-related coiled-coil, nucleotide-binding site, and leucine-rich repeat (CC-NB-LRR) encoding genes. However, only one gene conferred resistance to isolate Dg2, against which Rdg2a is effective, when the susceptible cultivar Golden Promise was transformed with the Rdg2a candidates. The high level of sequence similarity between the three genes most likely contributed to significant rearrangements during evolution, probably derived from unequal crossing-over resulting in sequence exchange between paralogs and in the generation of recombinant genes, as well as in expansion/contraction of gene copy number. To examine the haplotype variation at the Rdg2a locus, the sequencing of the allelic Mrdg2a (Morex rdg2a) locus of the Dg2-susceptible cultivar Morex was carried out and revealed large rearrangements including two deletions that generated an Rdg2a-homolog gene. This gene most likely derived from an un-equal crossing-over between the Rdg2a ancestor(s) and its paralog Nbs2-Rdg2a. PCR analyses performed with informative markers at eight loci within the Rdg2a locus identified eight different haplotypes. The Thibaut haplotype was observed to be largely conserved in Dg2-resistant barley cultivars. The resequencing of the Rdg2a gene in barley genotypes showing the same Thibaut haplotype or the same resistant phenotype revealed high sequence similarity to Thibaut Rdg2a, demonstrating the widespread conservation of the gene. Nonetheless, some sequence variation were identified in at least two barley genotypes that were verified for possible differences, with respect to Rdg2a, in the range of resistance specificities towards different leaf stripe isolates.

## Poster 60. Identification of a novel major leaf rust resistance QTL in the Swiss winter wheat cultivar Forno.

J. Singla <sup>1</sup>, S.G. Krattinger <sup>1,2</sup>, T. Wicker <sup>1</sup>, and B. Keller <sup>1</sup>.

<sup>1</sup> Institute of Plant Biology, University of Zurich, Zollikerstrasse 107, 8008 Zurich, Switzerland, and <sup>2</sup> CSIRO Plant Industry, GPO Box 1600, Canberra, ACT, 2601, Australia.

Leaf rust ( $Puccinia\ triticina$ ) is a major wheat disease which can cause significant yield losses globally. Rapidly evolving pathogen races increase the demand for more durable and race-nonspecific resistance genes. So far only four (Lr34, Lr46, Lr67, and Lr68) such durable leaf rust resistance genes have been identified. The identification and characterization of new sources of durable leaf rust resistance is important to ensure food security in the future. We earlier identified two major leaf rust resistance loci on chromosomes 1BS and 7DS in the Swiss winter wheat cultivar Forno. The resistance on chromosome 7DS was conferred by a single gene named Lr34 that encodes ABC transporter gene.

In this study we further characterized the second resistance locus on chromosome 1BS, named *QLrP.sfr-1BS*. This locus initially explained 28–32% of the phenotypic variance using 240 recombinant inbred lines (RILs) of the cross 'Arina/Frono'. We grouped these 240 RILs into the following categories: +*QLrP.sfr-1BS*/-*Lr34*, -*QLrP.sfr-1BS*/+*Lr34*, +*QLrP.sfr-1BS*/+*Lr34*, -*QLrP.sfr-1BS*/-*Lr34*, each having 21, 26, 22, and 18, lines respectively. The selected lines were phenotyped for the three traits, area under disease progress curve for the percentage of leaf area infected infection type, and leaf tip necrosis (LTN). The average length of LTN of only the *QLrP.sfr-1BS* group was significantly lower than the group having *Lr34*, indicating that unlike *Lr34*, *QLrP.sfr-1BS* is not associated with LTN. The groups only containing either *QLrP.sfr-1BS* or *Lr34* showed the same level of resistance, whereas the group containing both *QLrP.sfr-1BS* and *Lr34* show significantly higher level of resistance, demonstrating an additive effect of the two genes. A genetic linkage map for chromosome 1BS spanning a distance of 190 cM was constructed using SSRs and RFLP markers. Composite interval mapping detected *QLrP.sfr-1BS* with a LOD score of 17, flanked by markers wmc230 and gwm18. By adding